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Full Manual (EN)

EcoQprep™ Universal RNA Kit

- 50 rxn

K-3706

Name	Volume	Quantity	Name	Volume	Quantity
RD Buffer	30 mL	1 ea	1.5 mL Tube	50 ea	1 pack
RWMB1 Buffer	25 mL	1 ea	One Page Protocol	-	1 ea
RWB2 Buffer	15 mL	1 ea			
WE Buffer	40 mL	1 ea			
ER Buffer	25 mL	1 ea			
Magnetic Nanobead	6 mL	1 ea			

EcoQprep™ Universal RNA Kit

One Page Protocol

Before Start

Components	Before Use	Storage	Symbol
RD Buffer	Before each experiment, mix RD buffer with 10 µL of β-mercaptoethanol per 1 mL of RD buffer.	-	
RWMB1 and RWB2 Buffer	As indicated on the bottle label, add the correct amount of absolute ethanol (not provided).	Room temperature	
80% EtOH (Not provided)	80% EtOH is needed only for the drying bead method.	Room temperature	Run centrifuge Heating Add the following solution Caution

Protocol

NOTE: It is recommended to vortex after adding the solution.

Sample	Pre-treatment ①	Lysis ②	Precipitation ③	Binding ④	1 st Wash ⑤	2 nd Wash ⑥	3 rd Wash ⑦	Elution ⑧
Cultured Cells	Suspension cell culture Cell pellet ~1x10 ⁶ cells 300 xg, 5 min	+ 500 µL RD	+ 300 µL EtOH	+ 100 µL Bead 1) Attach magnet	+ 700 µL RWMB1 1) Attach magnet	+ 700 µL RWB2 1) Attach magnet	Perform either A or B A) Washing Bead + 700 µL WE ① Add to the Opposite side of the beads 1) Gently invert 2X 2) Discard sup 3) Detach magnet For 1) ~ 3), refer to 1 st and 2 nd Wash	+ 50 - 100 µL ER 60°C, 1 min 1) Attach magnet
	Monolayer cell culture Direct harvest 1) Discard cell culture medium			2) Invert 3~4X	2) Invert 3~4X	2) Invert 3~4X	B) Drying Bead + 700 µL 80% EtOH 1) Detach magnet 2) Attach magnet 3) Invert 3~4X 4) Discard sup 5) Detach magnet 6) 60°C, 5 min For 2) ~ 5), refer to 1 st and 2 nd Wash	2) Transfer the eluent
Plant tissue	Tissue : ~100 mg 1) Grind in LN ₂	+ 500 µL RD 60°C, 1-3 min full speed, 2 min Transfer the supernatant		3) Discard sup	3) Discard sup	3) Discard sup	Do Not Detach magnet	
	Animal tissue Homogenize sample : 20 ~ 30 mg	+ 500 µL RD full speed, 3 min Transfer the supernatant		4) Detach magnet	4) Detach magnet			